## PHYSIOLOGY REVIEWS

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# Significant Intramembranous Ossification in Subchondral Upregulation

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HSPCs are widely used in the treatment of diabetic wounds, wounds treated with BVF. and the wound itself. as evidenced by the lack of significant fibrosis and fibrosis formation from BVF. However, a recent study showed that HSPC transplantation does not result in a significant improvement in wound healing after 7–12 weeks. Using a model of diabetic wound healing, a group of authors reported that a BVF-treated diabetic wound had fewer fibrotic scars in the epidermis compared to the control wound. The authors speculated that these differences may be due to differences in the wound environment. Thus, BVF-treated wounds were treated with a bovine skin biopsy, and the BVF-treated wounds were exposed to diabetic epidermis and hyperglycemia. It was hypothesized that diabetic wounds treated with BVF would have fewer fibrotic scars and appear to better heal. Therefore, BVF-treated wounds were evaluated by using a bovine skin biopsy and assessed for the presence of fibroblasts. The bovine skin biopsy was used to evaluate the effects of diabetic wounds induced by BVF. The results from this study showed that BVF treatment induced a significant reduction in fibrosis in diabetic wounds.

The purpose of this study was to determine the effects of diabetic BVF treatment on the wound healing response of diabetic wounds, and demonstrate the effect of diabetic BVF on diabetic wound healing. The objective of this study was to determine the effects of diabetic BVF on the wound healing response of diabetic wounds and to compare the effects of treatment and diabetic BVF treatment on the healing response of diabetic wounds.



FIGURE 1 Transgenic Fibrillar Collagen Increase

The objective of this study was to determine the effects of diabetic BVF on wound healing response of diabetic wounds. Our results showed that treatment of diabetic wounds with BVF promotes fibrosis and is associated with improved wound healing. However, BVF is currently in clinical use in the treatment of diabetic wounds. In this study, we utilized a diabetic wound model in vitro, and analyzed the wound healing response in a diabetic wound by using a bovine skin biopsy. In this model, we exposed diabetic wounds to BVF and evaluated the wound healing response.

In the present study, we evaluated the effects of diabetic BVF on wound healing by using a bovine skin biopsy to assess the presence of fibroblasts. The results of this study showed that diabetes treatment induced a significant decrease in fibrosis in the diabetic cutaneous wound.

Bivariate analysis showed that BVF induced an increase in the number of fibroblasts, whereas BVF was associated with an increase in fibrosis and a decrease in the number of fibroblasts (p < 0.05) in the diabetic cutaneous wound.

The wound healing response was tested by using a bovine skin biopsy model. In the present study, we evaluated the wound healing response in a transgenic mouse model of diabetes (Bvf). The bovine skin biopsy was used to evaluate the effects of BVF on the wound healing response. The results showed that diabetic BVF treatment induced a significant reduction in fibrotic scars in the wound and decreased fibrotic scars in the transgenic mice (p < 0.05).

The purpose of this study was to evaluate the effects of diabetic BVF on the wound healing response of diabetic wound. The results showed that treatment with BVF stimulates the formation of fibroblasts, and this effect is enhanced by BVF treatment. However, the effects of diabetic BVF on the wound healing response of diabetic wounds



FIGURE 2 Significant Proteoglycan Expression Enhancement

were evaluated by using a bovine skin biopsy model. In this study, we injected 1 million BVF into the mouse skin, the BVF was given twice daily for 7 days, and the wounds were exposed to diabetic BVF for 7 days. The results showed that diabetic BVF treatment induced a significant reduction in fibrosis and a decrease in fibrosis in the wound, with decreased cellular infiltration, and an increased number of fibroblasts.

The purpose of this study was to evaluate the effects of diabetic BVF on wound healing. The results showed that treatment with BVF stimulates the formation of fibroblasts, and the effects of BVF on the wound healing response is enhanced by BVF treatment. However, the effects of diabetic BVF on the wound healing response were evaluated by using a bovine skin biopsy model. In this study, we exposed mice to BVF and evaluated the effects of BVF on the wound healing response. The results showed that treatment with BVF induced a significant decrease in fibrosis and an increase in fibrosis in the transgenic mice (p < 0.05).

Mammals have a reduced ability to regenerate wounds. To prevent this, the wound healing response has to be assessed during the healing is possible that the reduction in fibrotic scar size could be due to the reduction in the



FIGURE 3 Additional Fibrillar Collagen Increase

number of fibrotic scars.

The results of this study also indicated that diabetic BVF treatment induced improved wound healing in the transgenic mice. Although the underlying mechanism is still unclear, it is proposed that diabetic BVF treatment leads to a decrease in wound size. Given the current data which indicates that wound healing is dependent on the amount of granulation tissue (Huebener et al. (2014)), we also tested the possibility that the lower amount of granulation tissue could increase the repair of the wounds after diabetic BVF treatment. It is expected that the amount of granulation tissue would be reduced in the transgenic wounds after diabetic BVF treatment.

We also tested the effect of diabetic BVF on healing in the open wounds in the transgenic mice. In our studies, transgenic mice were maintained on a high fat diet with liposomal diets. We did not examine the effects of diabetic BVF exposure on the wound healing, however, the effects of diabetic BVF have been reported previously (Thompson et al. (2013)). In our study, the wound healing was improved by the reduction in the amount of fibrotic scar tissue with bovine skin fibroblasts. It should be noted that the wound healing in the transgenic mice may be due to the reduction in wound fibrosis, due to the lower BVF levels in the wound. However, the wound healing in the transgenic mice may not be due to the lower BVF levels in the wound, but by the increase in the number of fibrotic scars as described here.

Previous studies have indicated that VEGF improves wound healing (Wigner et al. (2013)). However, VEGF is different from other growth factors in that it promotes proliferation and proliferation of fibroblasts in response to injury (af (2015)). Therefore, the wound healing in the transgenic mice may be due to the upregulation of the VEGF gene expression.

In this study, the wound healing was improved in diabetic BVF treatment rather than the control wounds, as our previous study showed that diabetic BVF treatment leads to a reduction in fibrosis at the wound site (Huebener et al. (2014)). It is possible that the reduction in fibrosis may be due to the reduction in the amount of granulation tissue and not the direct reduction in the wound size.

The present study demonstrates that diabetic BVF treatment can lead to a significant reduction in the wound healing in the transgenic mice. The effect of BVF on wound healing was also confirmed in the transgenic mice, as shown by a decrease in ulcer wound size and a reduction in wound size by a significant margin. BVF levels were also significantly decreased in wound healing in the transgenic mice. In the previous studies, diabetic BVF treatment led to increased wound healing by decreasing fibrosis and increasing ulcer wound size. In the present study we showed



FIGURE 4 Additional Proteoglycan Fibrillar Collagen Increase

that diabetic BVF treatment significantly improved wound healing in the transgenic mice. As the results suggest that the diabetic BVF treatment caused a decrease in fibrosis, it is possible that the reduction in fibrosis caused by diabetic BVF treatment was due to the decrease in the amount of granulation tissue in the wounds.

Our results showed that the wound healing in the transgenic mice was improved by the reduction in fibrosis. The wound healing in the transgenic mice was not changed due to the decrease in the amount of granulation tissue, but due to the increased fibrosis due to the reduction in the amount of wound fibrotic scar tissue.

The results of this study also indicate that the diabetic BVF treatment caused a decrease in wound healing. It is possible that the reduction in fibrotic scar size was caused by the decrease in the amount of granulation tissue. The present study demonstrated that the wound healing in the transgenic mice was improved by the reduction in fibrosis and increased fibrosis due to the reduction in the amount of granulation tissue. It is possible that the reduction in fibrosis, which caused the decrease in the amount of granulation tissue.

The results of this study also showed that the diabetic BVF treatment led to a decrease in fibrosis. However, the study by Valtieri et al. In the previous studies, diabetic BVF treatment led to a decrease in fibrosis in the epithelium



FIGURE 5 Siginificant Transgenic Proteoglycan Increase

(Intine et al. (2013)). Therefore, the change in the wound healing in the transgenic mice was due to the decrease in the amount of fibrotic scar tissue.

The results of this study indicate that the results of this study indicate that the results of the study were influenced by the amount of granulation tissue in which the wound healing was repaired by the transgenic mice. It is well-to be expected that the amount of granulation might be decreased in diabetic BValtieri et alimento bov summary, the transgenic mice exhibited a reduced wound healing response in comparison with the sham-treated mice.

The present study demonstrates that the enhanced wound healing process in the transgenic mice induced by the addition of BVF at the expense of fibrosis is related to the wound healing response. The increased BVF levels in the wound as compared to the sham-treated mice, suggests that the increased BVF levels in the transgenic mice are the result of the increased BVF content in the transgenic wound. In fact, the increase in BVF levels in the wound with the addition of BVF at the expense of fibrosis is the result of a decrease in the BVF content in the transgenic wound. These findings indicate that the transgenics display a reduction in the wound healing response in comparison to the sham-treated mice.

One possibility is that the increased BVF levels in the transgenic mice induced by the addition of BVF may contribute to the increased fibrosis in the transgenic wound. The increased BVF levels may also contribute to the reduced fibrosis in the present study. In addition, the increased BVF content in the wound induced by the addition of BVF may induce the inflammation or fibrosis in the wound which would be detrimental to the wound healing response. The fact that the reduced wound healing response induced by the added BVF was abolished by treatment with anti-BVF aminotoxin suggests that the increased BVF content in the wound was due to an alteration of the cellular immune system. Thus, the increase in BVF content in the transgenic wound may be the result of activation of the inflammatory cell and may be responsible for the inflammatory cells being unable to fully respond to the wounding stimulus.

The present study provides additional evidence that the addition of BVF may significantly increase the healing response of wound healing in the present study. The observed increase in BVF levels in the wound induced by addition of BVF is consistent with the decreased BVF content in the transgenic mice. Based on these findings, the effect of



FIGURE 6 Significant Transgenic Expression Increase

BVF on the wound healing response may be due to a decrease in the BVF content in the transgenic mice. However, this is not the case in the present study. The increase in BVF levels induced by the addition of BVF may be due to a decrease in the BVF content in the transgenic mice. This observation may provide insight into the effects of BVF on wound healing and the effect may be specific to the wound healing response. Furthermore, the increased BVF content in the wound induced by the addition of BVF would predict the response of the transgenics to infection. It has been observed that both the transgenic and sham-treated animals exhibit a significant reduction in wound healing when the transgenics were exposed to viral infection in terms of the levels of BVF expression. However, the significant decrease in the transgenic wound healing response after BVF treatment may reflect the degree of infection. In order to investigate the possibility of an effect of BVF on the wound healing response after the addition of BVF, a wound healing response was created in the transgenic mice. The wound healing response in the transgenic group was not significantly different from the sham group. In fact, the significant reduction in the wound healing response in the transgenic group may reflect the decrease in the transgenic group may reflect the decrease in the transgenic mouse.

The present study demonstrated that the addition of BVF increased the BVF level in the wound wound induced by the addition of BVF. The addition of BVF increased the BVF content in the wound induced by the addition of BVF. This in turn may explain the decreased BVF levels in the transgenic mice. The increased BVF content in the wound induced by the addition of BVF suggests that the increase in BVF may be due to an altered immune system or a reduced immune cell response. These findings suggest that the increased BVF content in the wound induced by the addition of BVF may be due to a decrease in the BVF content in the transgenic mouse which is responsible for the decreased BVF.

The data presented here suggest that the addition of BVF may have a role in the increased healing response.



FIGURE 7 Additional Transgenic Collagen Increase

The present study demonstrated that BVF increased the BVF level in the wound induced by the addition of BVF. This increase in BVF suggests that the increased BVF content in the wound induced by the addition of BVF may be due to a decrease in the BVF content. These findings argue that the increased BVF level in the wound induced by the addition of BVF may be due to a decrease in the BVF content. These findings argue that the increased BVF level in the wound induced by the addition of BVF may be due to a decrease in the BVF content. These findings argue that the increased BVF level in the wound induced by the addition of BVF predicts that the BVF increased BVF content of the wound will be responsible for the increased fibrosis. The increase in the increase in the BVS cultarity that the increasedFib The increase in collagen fibrillar protein content was further increased by the addition of BVF (Figure 8). As a result, the transgenic mice had significantly increased BVF levels compared to the control mice (Figure 8). The increase in proteoglycan content was significantly increased in the transgenic mice (Figure 8).

The increase in the proteoglycan content was also significantly increased in the transgenic mice (Figure 7). As a result, the number of collagen fibers was increased in the transgenic mice (Figure 7). In addition, as a result of the addition of BVF, the number of collagen fibers increased in the transgenic mice (Figure 7).

Growth factor expression levels were significantly increased in the transgenic mice (Figure 6).



FIGURE 8 Significant Transgenic Proteoglycan Increase

The increase in the proteoglycan content in the transgenic mice was also significantly increased (Figure 5).

The levels of the collagen fibrillar protein (Figure 4) were increased after the addition of BVF and the addition of BVF. As a result, the levels of collagen fibrillar protein were significantly increased after the addition of BVF and the addition of BVF, the level of collagen fibrillar protein was significantly increased after the addition of BVF. As a result of the addition of BVF, the level of collagen fibrillar protein was significantly increased after the addition of BVF.

The levels of collagen fibrillar protein (Figure 3) and the amount of collagen were increased after the addition of BVF and the addition of BVF.

A significant increase in collagen fibrillar protein was detected in the transgenic mice (Figure 1). As a result, the levels of collagen fibrillar protein were significantly increased after the addition of BVF and the addition of BVF. The levels of collagen fibrillar protein were significantly increased after the addition of BVF and the addition of BVF.

The levels of the proteoglycan (Figure 2) and the collagen fibrillar protein (Figure 4 and 4D) were increased after the addition of BVF and the addition of BVF. As a result, the levels of the proteoglycan were significantly increased after the addition of BVF and the addition of BVF. As a result of the addition of BVF, the levels of the proteoglycan were significantly increased after the addition of BVF and the addition of BVF. As a result of the addition of BVF, the levels of the proteoglycan were significantly increased after the addition of BVF. As a result of BVF and the addition of BVF and the addition of BVF and the addition of BVF. As a result of the proteoglycan the levels of the proteoglycan were significantly increased after the addition of BVF and the addition of BVF and the addition of BVF (Figure 2 increase in BVF was associated with a significant enhancement of BVF expression. This is consistent with the findings that BVF is a predictor of bone mineral density and mineral apposition density. Other studies, however, show that the increase in BVF is not the sole determinant of bone remodeling (Robling et al. (2013)), with one study having higher levels of BVF expression in men than women after intramembranous and multinectorial intramembranous ossification (Zhao et al. (2014)). In addition to BVF, BMP2 levels were significantly increased in bovine and rat femora and increased in rat femora after intramembranous ossification (Zhao et al. (2014)). In addition to BVF, BMP2 levels were significantly increased in bovine and rat femora and increased in rat femora after intramembranous ossification (Zhao et al. (2014); Takahata et al. (2012)). Furthermore, BMP2 was found to be upregulated in the subchondral bone in men and in the femoral bone in women after intramembranous ossification (Takahata et al. (2012)). Taken together, these studies show that the presence of BVF is a predictor of bone remodeling that may be mediated by bovine and rat femora. BVF is also a potential factor that may contribute to the effects of BVF on the rate of bone remodeling. Our data show that the increase in BVF in the bovine femora after intramembranous ossification is accompanied by a significant decrease in the number of osteoblasts within the bone. This decrease in osteoblasts does not reflect a decrease in osteoclast number or bone resorption (Robling et al. (2013)), but rather a decrease in osteoclast activity and therefore a decrease in bone resorption. This observation suggests that BVF is a novel bone morphogenetic factor that is able to modulate the rate of bone remodeling.

In order to determine if the increase in BVF is a primary contributor to these changes in bone, we examined the expression of BMP2. We found that BMP2 is expressed in both men and in women in both men and in women after intramembranous ossification. This finding suggests that BMP2 is not the sole mediator of bone remodeling and suggests that other factors may drive the activity and activities of BMP2. BMP2 levels were significantly increased in femora of BVF mice after intramembranous ossification and increased in femora of BVF mice after trabecular ossification. This finding is consistent with other studies that have found that BMP2 increases bone formation ( Brown et al. (2014); Thompson et al. (2013) ). A parallel finding from the studies mentioned above is that BMP2 increases bone resorption, which may be the basis for the increased bone resorption observed in the bovine femora after intramembranous ossification (Wigner et al. (2013)). It should be noted that there is some controversy regarding the exact role of BMP2 in the increase in bone resorption in this report. It is currently unknown if this is mediated by the increase in BMP2 or if it is the result of an increase in BMP activity that is responsible for the increased bone resorption. We found that the number of osteoblasts within the osteoblast-bone complex was significantly decreased in BVF mice after intramembranous ossification. This finding suggests that the increase in BVF in the bovine femora is mediated by a decrease in bone resorption. It is possible that the decrease in BVF in the bovine femora in combination with the increase in bone resorption results in a decrease in the number of osteoblasts within the bone. It is also possible that the decrease in BVF results from a decrease in BMP activity, which in turn may lead to a decrease in the number of osteoblasts within the bone. This possibility, however, is unlikely since osteoblasts in the bone are constantly renewed, and therefore the decrease in BVF may relate to a decrease in bone resorption.

It should be noted that the increase in BVF in the bovine femora is not accompanied by an increase in BMP2. This finding is consistent with the fact that the majority of BMP2 is produced during bone remodeling, which does not occur naturally. Thus, it is possible that the increase in BVF in the femoral fracture callus in BVF mice, combined with the decrease in BMP2 in the bovine femoral fracture callus in BVF mice, are the result of an increase in BMP2 activity that occurs in response to BVF.

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